1,2,4,8,13,14,20,22,24,28,32,46

3/7/1 (Item 1 from file: 5)
DIALOG(R)File 5:BIOSIS PREVIEWS(R)

11807415 BIOSIS Number: 98407415
Growth of Donor-derived Dendritic Cells from the Bone Marrow of Murine Liver Allograft Recipients in Response to Granulocyte-Macrophage Colony-stimulating Factor
Lu L; Rudert W A; Qian S; McCaslin D; Fu F; Rao A S; Trucco M; Fung J J; Starzl T E; Thomson A W
Dep. Surg., Univ. Pittsburgh Med. Cent., W1544 Biomed. Sci. Tower, Terrace and Lothrop St., Pittsburgh, PA 15213, USA
Journal of Experimental Medicine 182 (2). 1995. 379-387.
Full Journal Title: Journal of Experimental Medicine
ISSN: 0022-1007
Language: ENGLISH
Print Number: Biological Abstracts Vol. 100 Iss. 006 Ref. 085007
Allografts of the liver, which has a comparatively heavy leukocyte content compared with other vascularized organs, are accepted permanently across major histocompatibility complex barriers in many murine strain combinations without immunosuppressive therapy. It has been postulated that this inherent tolerogenicity of the liver may be a consequence of the migration and perpetuation within host lymphoid tissues of potentially tolerogenic donor-derived ("chimeric") leukocytes, in particular, the precursors of chimeric dendritic cells (DC). In this study, we have used granulocyte/macrophage colony-stimulating factor to induce the propagation of progenitors that give rise to DC (CD45+, CD11c+, 33D1+, nonlymphoid dendritic cell 145+, major histocompatibility complex class II+, B7-1+) in liquid cultures of murine bone marrow cells. Using this technique, together with immunocytochemical and molecular methods, we show that, in addition to cells expressing female host (C3H) phenotype (H-2K-k+; I-E+; Y chromosome-), a minor population of male donor (B10)-derived cells (H-2K-b+; I-A+; Y chromosome+) can also be grown in 10-d DC cultures from the bone marrow of liver allograft recipients 14 d after transplant. Highly purified nonlymphoid dendritic cell 145+ DC sorted from these bone marrow-derived cell cultures were shown to comprise apprx 1-10% cells of donor origin (Y chromosome+) by polymerase chain reaction analysis. In addition, sorted DC stimulated naive, recipient strain T lymphocytes in primary mixed leukocyte cultures. Evidence was also obtained for the growth of donor-derived cells from the spleen but not the thymus. In contrast, donor cells could not be propagated from the bone marrow or other lymphoid tissues of nonimmunosuppressed C3H mice rejecting cardiac allografts from the same donor strain (B10). These findings provide a basis for the establishment and perpetuation of cell chimerism after organ transplantation.

3/7/2 (Item 2 from file: 5)
DIALOG(R)File 5:BIOSIS PREVIEWS(R)

11789070 BIOSIS Number: 98389070
Isolation of a Small Monocyte Subset With High Accessory Function: A Dendritic Cell Precursor?
Uren S; Boyle W
Dep. Microbiol., Univ. Melbourne, Gratten St., Parkville, VIC 3052, Australia
Transplantation Proceedings 27 (3). 1995. 2172-2173.
Full Journal Title: Twelfth Scientific Meeting of the Transplantation Society of Australia and New Zealand, April 20-22, 1994. Transplantation

Proceedings
ISSN: 0041-1345
Language: ENGLISH
Print Number: Biological Abstracts/RRM Vol. 047 Iss. 009 Ref. 151473

3/7/4 (Item 4 from file: 5)
DIALOG(R)File 5:BIOSIS PREVIEWS(R)

11723693 BIOSIS Number: 98323693
Ultrastructural and immunophenotypic differentiation of dendritic cells from mouse bone marrow cultures supplemented with granulocyte-macrophage colony-stimulating factor (GM-CSF)
Umezu H; Naito M; Takashashi K I A
Second Dep. Pathol., Niigata Univ. Sch. Med., 1 Asahimachi-Dori, Niigata 951, Japan
Journal of Submicroscopic Cytology and Pathology 27 (2). 1995. 227-234.
Full Journal Title: Journal of Submicroscopic Cytology and Pathology
ISSN: 1122-9497
Language: ENGLISH
Print Number: Biological Abstracts Vol. 100 Iss. 003 Ref. 031417
The present study examined electron microscopical, immunohistochemical and morphometric changes in the development, differentiation, and maturation of dendritic cells in the culture of mouse bone marrow cells supplemented with granulocyte/macrophage colony-stimulating factor (GM-CSF). A large number of large dendritic cells were released from the aggregates of small round cells. These released cells showed morphological, immunophenotypic, and functional characteristics of typical dendritic cells. These dendritic cells possessed irregularly-shaped nuclei in abundant cytoplasm with rough endoplasmic reticulum (rER), vesicles, multivesicular bodies, and tubulovesicular system, and projected long dendritic cytoplasmic processes. The tubulovesicular system and dendritic surface projections were characteristic of interdigitating cells usually residing in the paracortical area of lymph nodes. Small cells in aggregates were round in shape, had oval nuclei in narrow cytoplasm with poorly developed intracellular organelles, and projected a few short processes. These cells were a proliferating population distinct from monocytes in both ultrastructure and immunophenotype. Morphometrical analysis of cultured cells provided evidence that the small cells differentiate into typical dendritic cells via intermediate cells. These results imply that dendritic cells differentiate from small round dendritic precursor cells and that GM-CSF is a major cytokine capable of inducing the development, differentiation, and proliferation of dendritic cells.

3/7/8 (Item 8 from file: 5)
DIALOG(R)File 5:BIOSIS PREVIEWS(R)

11228245 BIOSIS Number: 97428245
Isolation and function of human dendritic cells
Williams L A; Egner W; Hart D N J
Haematol./Immunol. Res. Group, Christchurch Hosp., Christchurch, NEZ 0 (0). 1994. 41-103.
Full Journal Title: Jeon, K. W. and J. Jarvik (Ed.). International Review of Cytology, Vol. 153. x+305p. Academic Press, Inc.: San Diego, California, USA; London, England, UK. ISBN 0-12-364556-5.
ISSN: 0074-7696
Language: ENGLISH
Print Number: Biological Abstracts/RRM Vol. 046 Iss. 010 Ref. 153009

3/7/13 (Item 13 from file: 5)
DIALOG(R)File 5:BIOSIS PREVIEWS(R)

10004121 BIOSIS Number: 95004121
CYTOKINE GENE EXPRESSION IN MURINE EPIDERMAL CELL SUSPENSIONS INTERLEUKIN 1-BETA AND MACROPHAGE INFLAMMATORY PROTEIN 1-ALPHA ARE SELECTIVELY EXPRESSED IN LANGERHANS CELLS BUT ARE DIFFERENTIALLY REGULATED IN CULTURE
HEUFLER C; TOPAR G; KOCH F; TROCKENBACHER B; KAMPGEN E; ROMANI N; SCHULER G
DEP. DERMATOL., UNIV. INNSBRUCK, ANICHSTR. 35, INNSBRUCK A-6020, AUSTRIA.
J EXP MED 176 (4). 1992. 1221-1226. CODEN: JEMEA
Full Journal Title: Journal of Experimental Medicine
Language: ENGLISH
Epidermal Langerhans cells (LC) are considered direct yet immature precursors of dendritic cells (DC) in the draining lymph nodes. Although the development of LC into potent immunostimulatory DC occurs in vitro and has been studied in detail, little is known about their profile of cytokine gene expression. By using reverse transcriptase polymerase chain reaction analysis to screen 16 cytokines followed by Northern blotting for selected analysis, we determined the cytokine gene expression profile of murine LC at different time points in culture when T cell stimulatory activity is increasing profoundly. LC regularly expressed macrophage inflammatory proteins, MIP-1.alpha. and MIP-2, and interleukin 1.beta. (IL-1.beta.). Both MIPs were downregulated upon culture and maturation into DC, whereas IL-1.beta. was strongly upregulated in culture. MIP-1.alpha. and IL-1.beta. mRNA were found only in LC, but not in other epidermal cells. Apart from trace amounts of IL-6 in cultured LC, several macrophage and T cell products were not detected. The cytokine expression profile of LC thus appears distinct from typical macrophages. The exact role of the cytokine genes we found transcribed in LC remains to be determined.

3/7/14 (Item 14 from file: 5)
DIALOG(R)File 5:BIOSIS PREVIEWS(R)

9512505 BIOSIS Number: 94017505
IDENTIFICATION OF PROLIFERATING DENDRITIC CELL PRECURSORS IN MOUSE BLOOD
INABA K; STEINMAN R M; PACK M W; AYA H; INABA M; SUDO T; WOLPE S; SCHULER G
LAB. CELL. PHYSIOL. IMMUNOL, BOX 280, ROCKEFELLER UNIV., 1230 YORK AVE., NEW YORK, N.Y. 10021.
J EXP MED 175 (5). 1992. 1157-1167. CODEN: JEMEA
Full Journal Title: Journal of Experimental Medicine
Language: ENGLISH
While it has been know that dendritic cells arise from proliferating precursors in situ, it has been difficult to identify progenitors in culture. We find that aggregates of growing dendritic cells develop in cultures of mouse blood that are supplemented with granulocyte/macrophage colony-stimulating factor (GM-CSF) but not other CSFs. The dendritic cell precursor derives from the Ia-negative and nonadherent fraction. The aggregates of developing dendritic cells appear at about 1 wk of culture, with 100 · or more such clusters being formed per 106 blood leukocytes. The aggregates can be dislodged and subcultured as expanding clusters that are covered with cells having the motile sheet-like processes ("veils") of dendritic cells. By about 2 wk, large numbers of single, major histocompatibility complex (MHC) class II-rich dendritic cells begin to be relased into the medium. Combined immunoperoxidase and [3H]thymidine autoradiography show that the cells that proliferate within the aggregate lack certain antigenic markers that are found on mature dendritic cells. However, in pulse-chase protocols, the [3H]thymidine-labeled progeny exhibit many typical dendritic cell features, including abundant MHC class II and a cytoplasmic granular antigen identified by monoclonal antibody 2A1. The progeny dendritic cells are potent stimulators of the mixed leukocyte reaction and can home to the T-dependent areas of lymph node after injection into the footpads. We conlcude that mouse blood contains GM-CSF-dependent, proliferating progenitors that give rrise to large numbers of dendritic cells with characteristic morphology, mobility,

3/7/20 (Item 20 from file: 5)
DIALOG(R)File 5:BIOSIS PREVIEWS(R)

8430842 BIOSIS Number: 41114842
THE ROLE OF FETAL EPITHELIAL TISSUES IN THE MATURATION-DIFFERENTIATION OF BONE MARROW-DERIVED PRECURSORS INTO DENDRITIC EPIDERMAL T CELLS DETC OF THE MOUSE
STINGL G; ELBE A; PAER E; KILGUS O; STROHAL R; SCHREIBER S
DIV. CUTANEOUS IMMUNOL., DEP. DERMATOL. I, UNIV. VIENNA MED. SCH., VIRCC, BRUNNER-STR. 59, A-1235 VIENNA, AUSTRIA.
PFEFFER, K., ET AL. (ED.). CURRENT TOPICS IN MICROBIOLOGY AND IMMUNOLOGY, VOL. 173. FUNCTION AND SPECIFICITY OF GAMMA/DELTA CELLS; INTERNATIONAL WORKSHOP, SCHLOSS ELMAU, BAVARIA, GERMANY, OCTOBER 14-16, 1990. XII+296P. SPRINGER-VERLAG: BERLIN, GERMANY; NEW YORK, NEW YORK, USA. ILLUS. ISBN 3-540-53781-3; ISBN 0-387-53781-3. 0 (0). 1991. 269-278. CODEN: CTMIA
Language: ENGLISH

3/7/22 (Item 22 from file: 5)
DIALOG(R)File 5:BIOSIS PREVIEWS(R)

7518933 BIOSIS Number: 39031540
ESTABLISHMENT OF A CELL LINE OF PUTATIVE CD3-POSITIVE PRECURSORS OF DENDRITIC EPIDERMAL T CELLS DETO FROM FETAL SKIN
ELBE A; KILGUS O; STROHAL R; SCHINHAN H; STINOL G
DIV. CUTANEOUS IMMUNOBIOL., DEP. DERMATOL. I, UNIV. VIENNA MED. SCH., VIRCC, SFI, BRUNNER STR. 59. A-1235 VIENNA, AUSTRIA.
JOINT MEETING OF THE AMERICAN SOCIETY FOR BIOCHEMISTRY AND MOLECULAR BIOLOGY AND THE AMERICAN ASSOCIATION OF IMMUNOLOGISTS, NEW ORLEANS, LOUISIANA, USA, JUNE 4-7, 1990. FASEB (FED AM SOC EXP BIOL) J 4 (7). 1990. A1736. CODEN: FAJOE
Language: ENGLISH

3/7/24 (Item 24 from file: 5)
DIALOG(R)File 5:BIOSIS PREVIEWS(R)

7353598 BIOSIS Number: 89004617
CULTURED HUMAN LANGERHANS CELLS RESEMBLE LYMPHOID DENDRITIC CELLS IN PHENOTYPE AND FUNCTION
ROMANI N; LENZ A; GLASSEL H; STOESSEL H; STANZL U; MAJDIC O; FRITSCH P; SCHULER G
DEP. DERMATOL., UNIV. INNSBRUCK, A-6020 INNSBRUCK, AUSTRIA.
J INVEST DERMATOL 93 (5). 1989. 600-609. CODEN: JIDEA
Full Journal Title: Journal of Investigative Dermatology
Language: ENGLISH
Freshly isolated murine epidermal Langerhans cells (LC) are weak stimulators of resting T cells. Upon culture their phenotype changes, their stimulatory activity increases significantly, and they come to resemble lymphoid dendritic cells. Resident murine LC, therefore, might represent a reservoir of immature dendritic cells. We have now used enzyme cytochemistry, a panel of some 80 monoclonal antibodies, and immunofluorescence microscopy or two-color flow cytometry, as well as transmission electron microscopy, to analyse the phenotype and morphology of human LC before and after 2-4 d of bulk epidermal cell culture. In addition, LC were enriched from bulk epidermal cell cultures, and their stimulatory capacity was tested in the allogeneic mixed leukocyte reaction and the oxidative mitogenesis assay. Cultured human LC resembled human lymphoid dendritic cells in morphology, phenotype, and function. Specifically, LC became non-adherent upon culture and developed sheet-like

activity, and lost nonspecific esterase activity. As in the mouse, surface expression of MHC class I and II antigens increased significantly, and FcII receptors were significantly reduced. Markers that are expressed by dendritic cells (like CD40) appeared on LC following culture. Cultured human LC were potent T-cell stimulators. Our findings support the view that resident human LC, like murine LC, represent immature precursors of lymphoid dendritic cells in skin-draining lymph nodes.

3/7/28 (Item 28 from file: 5)
DIALOG(R)File 5:BIOSIS PREVIEWS(R)

5645308 BIOSIS Number: 33040329
PROPERTIES OF DENDRITIC CELL PRECURSORS IN RAT BONE MARROW
BOWERS W E; BERKOWITZ M R
BASSETT RES. INST. MED. RES., COPPERSTOWN, N.Y. 13326.
SYMPOSIUM ON RECENT ADVANCES IN LEUKEMIA AND LYMPHOMA HELD AT THE 16TH ANNUAL MEETING OF THE UCLA (UNIVERSITY OF CALIFORNIA-LOS ANGELES) SYMPOSIA ON MOLECULAR AND CELLULAR BIOLOGY, LOS ANGELES, CALIFORNIA, USA, JANUARY 25-31, 1987. J CELL BIOCHEM SUPPL 0 (11 PART A). 1987. 233. CODEN: JCBSD
Language: ENGLISH

3/7/32 (Item 32 from file: 5)
DIALOG(R)File 5:BIOSIS PREVIEWS(R)

4751317 BIOSIS Number: 29108632
DENDRITIC CELL PRECURSORS IN RAT BONE MARROW
BOWERS W E; BERKOWITZ M R
BASSETT INST. FOR MED. RES., COOPERSTOWN, NY 13326.
JOINT CONFERENCE OF THE 17TH INTERNATIONAL LEUKOCYTE CULTURE CONFERENCE AND THE 22ND NATIONAL MEETING OF THE RETICULOENDOTHELIAL SOCIETY, ITHACA, N.Y., USA, AUG. 3-8, 1985. J LEUKOCYTE BIOL 38 (1). 1985. 166. CODEN: JLBIE
Language: ENGLISH

3/7/46 (Item 8 from file: 155)
DIALOG(R)File 155:MEDLINE(R)

05619814 85235814
A comparison of murine epidermal Langerhans cells with spleen dendritic cells.
Schuler G; Romani N; Steinman RM
J Invest Dermatol (UNITED STATES) Jul 1985, 85 (1 Suppl) p99s-106s, ISSN 0022-202X Journal Code: IHZ
Contract/Grant No.: AI 13013
Languages: ENGLISH
Document type: JOURNAL ARTICLE
To establish if epidermal Langerhans cells (LC) are related to spleen dendritic cells, we have considered the morphology, phenotype, and function of the 2 cell types in culture. Cultured LC could be partially enriched (up to 50%) on the basis of 2 simple physical properties: nonadherence to plastic, and low buoyant density in dense albumin columns. The morphology of cultured LC and spleen dendritic cells were similar. In particular both cell types had many cell processes and/or veils, and cultured LC lost their distinguishing Birbeck granules. Freshly isolated LC exhibited nonspecific esterase and ATPase, as well as the F4/80 (alpha-macrophage) and 2.4G2 (alpha-Fc receptor) antigens. However all these traits were lost in culture, while Ia and Mac-1 antigens persisted. As a result, the cytochemical and antigenic phenotype of LC became similar to spleen dendritic cells. The one exception was that LC lacked the 33D1 dendritic

cells in that fresh LC were weak stimulators of T cell proliferation in the mixed leukocyte reaction and in sodium periodate-induced mitogenesis. However, stimulatory activity per cell increased at least 30 fold in culture so that by 2-3 days, LC were 3-10 times more potent than dendritic cells. Maturation of LC function was radioresistant and was accompanied by a small increase in cell surface Ia antigens. Although LC have been likened both to lymphoid dendritic cells and to macrophages, our data suggest a different conclusion. LC seem to be dendritic cell precursors and are immunologically immature. Possibly, lymphoid dendritic cells are in general derived from substantial pools of precursors in nonlymphoid tissues, such as epidermal LC.

?t s7/7/3,4,8,15

7/7/3 (Item 3 from file: 5)
DIALOG(R)File 5:BIOSIS PREVIEWS(R)

11807133 BIOSIS Number: 98407133
Dendritic Cells Use Macropinocytosis and the Mannose Receptor to Concentrate Macromolecules in the Major Histocompatibility Complex Class II Compartment: Downregulation by Cytokines and Bacterial Products

Sallusto F; Cella M; Danieli C; Lanzavecchia A
Basel Inst. Immunol., Grenzacherstrasse 487, CH-4005 Basel, Switzerland

Journal of Experimental Medicine 182 (2). 1995. 389-400.
Full Journal Title: Journal of Experimental Medicine
ISSN: 0022-1007
Language: ENGLISH
Print Number: Biological Abstracts Vol. 100 Iss. 006 Ref. 084725

We have previously demonstrated that human peripheral blood low density mononuclear cells cultured in granulocyte/macrophage colony-stimulating factor (GM-CSF) and interleukin (IL)-4 develop into dendritic cells (DCs) that are extremely efficient in presenting soluble antigens to T cells. To identify the mechanisms responsible for efficient antigen capture, we studied the endocytic capacity of DCs using fluorescein isothiocyanate-dextran, horseradish peroxidase, and lucifer yellow. We found that DCs use two distinct mechanisms for antigen capture. The first is a high level of fluid phase uptake via macropinocytosis. In contrast to what has been found with other cell types, macropinocytosis in DCs is constitutive and allows continuous internalization of large volumes of fluid. The second mechanism of capture is mediated via the mannose receptor (MR), which is expressed at high levels on DCs. At low ligand concentrations, the MR can deliver a large number of ligands to the cell in successive rounds. Thus, while macropinocytosis endows DCs with a high capacity, nonsaturable mechanism for capture of any soluble antigen, the MR gives an extra capacity for antigen capture with some degree of selectivity for non-self molecules. In addition to their high endocytic capacity, DCs from GM-CSF+IL-4-dependent cultures are characterized by the presence of a large intracellular compartment that contains high levels of class II molecules, cathepsin D, and lysosomal-associated membrane protein-1, and is rapidly accessible to endocytic markers. We investigated whether the capacity of DCs to capture and process antigen could be modulated by exogenous stimuli. We found that DCs respond to tumor necrosis factor alpha, CD40 ligand, IL-1, and lipopolysaccharide with a coordinate series of changes that include downregulation of macropinocytosis and Fc receptors, disappearance of the class II compartment, and upregulation of adhesion and costimulatory molecules. These changes occur within 1-2 d and are irreversible, since neither pinocytosis nor the class II compartment are recovered when the maturation-inducing stimulus is removed. The specificity of the MR and the capacity to respond to inflammatory stimuli maximize the capacity of DCs to present infectious non-self antigens to T cells.

7/7/4 (Item 4 from file: 5)
DIALOG(R)File 5:BIOSIS PREVIEWS(R)

11795192 BIOSIS Number: 98395192
Induction of dendritic morphology in murine B-cells by interleukin 4
Davey E J; Conrad D H; Severinson E
Nobel Inst. Cell Mol. Biol., Stockholm, Sweden
0 (0). 1995. 48.
Full Journal Title: 9TH INTERNATIONAL CONGRESS OF IMMUNOLOGY. The 9th International Congress of Immunology; Meeting Sponsored by the American Association of Immunologists and the International Union of Immunological Societies, San Francisco, California, USA, July 23-29, 1995. ix+742p. 9th International Congress of Immunology: San Francisco, California, USA.
ISSN: *********
Language: ENGLISH
Print Number: Biological Abstracts/RRM Vol. 047 Iss. 009 Ref. 157595

7/7/8 (Item 8 from file: 5)
DIALOG(R)File 5:BIOSIS PREVIEWS(R)

11755082 BIOSIS Number: 98355082
Interleukin-13 selectively suppresses the growth of human macrophage progenitors at the late stage
Sakamoto O; Hashiyama M; Minty A; Ando M; Suda T
Dep. Cell Differentiation, IMEG, Kumamoto Univ. Sch. Med., 2-2-1 Honjo, Kumamoto 860, Japan
Blood 85 (12). 1995. 3487-3493.
Full Journal Title: Blood
ISSN: 0006-4971
Language: ENGLISH
Print Number: Biological Abstracts Vol. 100 Iss. 004 Ref. 046920

Interleukin-13 (IL-13) is a pleiotropic cytokine that inhibits the production of inflammatory cytokines of monocytes. We investigated the effects of IL-13 on the clonal growth of human hematopoietic progenitors. IL-13 alone did not support any colony formation. IL-13 markedly suppressed macrophage colonies that were formed in the presence of IL-3 and erythropoietin, granulocyte-macrophage colony-stimulating factor, or macrophage colony-stimulating factor. Macrophage colony cells showed dendritic cell-like morphology and cellular aggregates. IL-13 did not affect granulocyte colony and erythroid burst formation. Delayed addition of IL-13 and replating onto the culture dishes with IL-13 showed that macrophage colony formation was suppressed during days 8 and 14 of culture. These results indicate that IL-13 affects the growth of the late stage of committed macrophage progenitors. Single-cell culture of isolated CD34+CD33+ cells with IL-13 confirmed that macrophage colony formation was significantly suppressed. These results show that IL-13 directly suppresses the proliferation of differentiating macrophages. In addition, these suppressive effects of IL-13 were synergistic with IL-4. Furthermore, in the liquid culture of bone marrow cells in the presence of IL-13, the number of CD14 (monocyte-macrophage antigen) positive cells decreased and CD18 (LFA-1-beta)-positive cells increased. It is concluded that IL-13 affects the growth of the late stage of macrophage precursors as well as mature monocytes. Induction of differentiation of human monocytes may be correlated with the suppression of their progenitors.

7/7/15 (Item 15 from file: 5)
DIALOG(R)File 5:BIOSIS PREVIEWS(R)

10902293 BIOSIS Number: 97102293
IL-4 stimulates the generation of dendritic cells and multinucleated giant cells from human monocytes
Akagawa K S; Takasuka N; Sakurai T
Dep. Immunol., Natl. Inst. Health, 1-23-1, Toyama, Shinjuku-ku, Tokyo

Lymphokine and Cytokine Research 12 (5). 1993. 326.
Full Journal Title: Combined Meeting of the 8th International Lymphokine Workshop and the 4th International Workshop on Cytokines: Lymphokines and Cytokines from Clone to Clinic, Osaka, Japan, October 17-21, 1993. Lymphokine and Cytokine Research
ISSN: 1056-5477
Language: ENGLISH
Print Number: Biological Abstracts/RRM Vol. 046 Iss. 003 Ref. 031186

? t s7/7/13,14

QR185.8.L93L95

7/7/13 (Item 13 from file: 5)
DIALOG(R)File 5:BIOSIS PREVIEWS(R)

11058686 BIOSIS Number: 97258686
Induction of a primary IFN-g response by Peyer's patch dendritic cells is mediated by IL-12 and inhibited by IL-4
Kelsall B L; Ehrhardt R O; Strober W
Lab. Clin. Investigation, Natl. Health, Bethesda, MD 20892, USA
FASEB Journal 8 (4-5). 1994. A207.
Full Journal Title: Experimental Biology 94, Parts I and II, Anaheim, California, USA, April 24-28, 1994. FASEB Journal
ISSN: 0892-6638
Language: ENGLISH
Print Number: Biological Abstracts/RRM Vol. 046 Iss. 006 Ref. 091866

7/7/14 (Item 14 from file: 5)
DIALOG(R)File 5:BIOSIS PREVIEWS(R)

R850.J6

11039158 BIOSIS Number: 97239158
Efficient presentation of soluble antigen by cultured human dendritic cells is maintained by granulocyte-macrophage colony-stimulating factor plus interleukin 4 and downregulated by tumor necrosis factor alpha
Sallusto F; Lanzavecchia A
Basel Inst. Immunol., Grenzacherstr. 487, CH-4005 Basel, SWI
Journal of Experimental Medicine 179 (4). 1994. 1109-1118.
Full Journal Title: Journal of Experimental Medicine
ISSN: 0022-1007
Language: ENGLISH
Print Number: Biological Abstracts Vol. 097 Iss. 011 Ref. 156271
Using granulocyte/macrophage colony-stimulating factor (GM-CSF) and interleukin 4 we have established dendritic cell (DC) lines from blood mononuclear cells that maintain the antigen capturing and processing capacity characteristic of immature dendritic cells in vivo. These cells have typical dendritic morphology, express high levels of major histocompatibility complex (MHC) class I and class II molecules, CD1, Fc-gamma-RII, CD40, B7, CD44, and ICAM-1, and lack CD14. Cultured DCs are highly stimulatory in mixed leukocyte reaction (MLR) and are also capable of triggering cord blood naive T cells. Most strikingly, these DCs are as efficient as antigen-specific B cells in presenting tetanus toxoid (TT) to specific T cell clones. Their efficiency of antigen presentation can be further enhanced by specific antibodies via FcR-mediated antigen uptake. Incubation of these cultured DCs with tumor necrosis factor alpha (TNF-alpha) or soluble CD40 ligand (CD40L) for 24 h results in an increased surface expression of MHC class I and class II molecules, B7, and ICAM-1 and in the appearance of the CD44 exon 9 splice variant (CD44-v9); by contrast, Fc-gamma-RII is markedly and sometimes completely downregulated. The functional consequences of the short contact with TNF-alpha are an increased T cell stimulatory capacity in MLR, but a 10-fold decrease in presentation of soluble TT and a 100-fold decrease in presentation of TT-immunoglobulin G comple[illegible].
?ds

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S1           0   DENDRITIC(2A)PRECURSOR?
S2          76   DENDRITIC([redacted](PRECURSOR OR PRECURSORS)
S3          50   RD S2 (uni[redacted] items)
S4       16168   (INTERLEUKIN OR IL)(2N)(4 OR 13)
S5        1622   S4(10N)(MACROPHAGE OR MACROPHAGES)
S6          45   S4(10N)DENDRITIC
S7          32   RD S6 (unique items)
?d
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1. 5,446,090, Aug. 29, 1995, Isolatable, water soluble, and hydrolytically stable active sulfones of poly(ethylene glycol) and related polymers for modification of surfaces and molecules; J. Milton Harris, 525/54.1; 424/94.3; 435/188; 514/2; 525/50, 54.11, 60, 409, 535; 528/374; 530/351, 404; 568/32, 621, 623 [IMAGE AVAILABLE]

=> d 1-11

1. 5,439,907, Aug. 8, 1995, Use of N9 morpholino derivatives of 7,8-disubstituted guanines; Robert Chen, et al., 514/234.2; 435/240.2, 240.25 [IMAGE AVAILABLE]

2. 5,422,110, Jun. 6, 1995, Enhanced immunogenicity using leukotoxin chimeras; Andrew A. Potter, et al., 424/255.1, 184.1, 190.1, 192.1, 234.1; 435/69.1, 69.3, 172.1, 172.3; 530/350, 825; 536/23.4, 23.7 [IMAGE AVAILABLE]

3. 5,417,986, May 23, 1995, Vaccines against diseases caused by enteropathogenic organisms using antigens encapsulated within biodegradable-biocompatible microspheres; Robert H. Reid, et al., 424/499, 422, 426, 433, 444, 455, 470, 486, 488, 489, 491 [IMAGE AVAILABLE]

4. 5,382,580, Jan. 17, 1995, N9 morpholino derivatives of 7,8-disubstituted guanines; Robert Chen, et al., 514/234.2; 544/118 [IMAGE AVAILABLE]

5. 5,354,686, Oct. 11, 1994, Extracellular matrix protein adherent T cells; Allan B. Haberman, 435/240.2, 240.23, 240.243 [IMAGE AVAILABLE]

6. 5,284,931, Feb. 8, 1994, Intercellular adhesion molecules, and their binding ligands; Timothy A. Springer, et al., 424/139.1, 152.1, 153.1, 154.1, 172.1, 173.1; 514/8; 530/388.22, 395, 808, 868 [IMAGE AVAILABLE]

7. 5,227,298, Jul. 13, 1993, Method for microencapsulation of cells or tissue; Collin J. Weber, et al., 435/178; 264/4.3, 4.32, 4.33; 424/424, 493, 497 [IMAGE AVAILABLE]

8. 5,206,345, Apr. 27, 1993, **IL**-**4** and TNF induce mAb 6G10-recognized expression on bone marrow stromal cells; Boris Masinovsky, et al., 530/388.7; 435/7.21, 240.27; 436/548 [IMAGE AVAILABLE]

9. 5,188,959, Feb. 23, 1993, Extracellular matrix protein adherent T cells; Allan B. Haberman, 435/240.243, 4, 240.2 [IMAGE AVAILABLE]

10. 5,106,731, Apr. 21, 1992, Kaposi's sarcoma endothelial cells and growth factor; Syed Z. Salahuddin, et al., 435/7.24, 7.23, 240.2, 804; 530/300, 351, 388.24, 389.2, 828 [IMAGE AVAILABLE]

11. 4,789,658, Dec. 6, 1988, Immunoprophylactic and immunotherapeutic agents; Ryota Yoshimoto, et al., 514/2; 435/69.52, 70.2, 70.3, 70.4; 514/8, 12; 530/351, 370, 371 [IMAGE AVAILABLE]

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(FILE 'USPAT' ENTERED AT 13:44:09 ON 12 OCT 93)

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L1             1 S DENDRITIC(2A)PRECURSOR?
L2          1986 S DENDRITIC
L3           469 S (INTERLEUKIN OR IL)(2A)(4 OR 13)
L4            11 S L2 AND L3

=>
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